Xenoestrogens and Metainflammation: An interplay between Immune System, Metabolism and Obesity

Santanu Basak, Archna Panghal and Gopabandhu Jena*

Facility for Risk Assessment and Intervention Studies, Dept. of Pharmacology and Toxicology, National Institute of Pharmaceutical Education and Research, S.A.S Nagar, Punjab, India, 160062 gbjena@niper.ac.in

Obesity is the greatest threat to mankind in twenty first century. It is accompanied with low grade chronic inflammation (metainflammation) which persists in all the tissues involved in energy balance. Obesogens such as xenoestrogens enter into the body and disrupt normal metabolism of cells which are responsible for fat deposition namely adipocytes. The research findings demonstrated that macrophages play a crucial role in metabolic tissues during the onset of obesity. When an intricate balance between metabolism and innate immunity is disturbed, macrophages surrounding adipocytes get activated and converted into pro-inflammatory subtypes. Pro-inflammatory macrophages secret pro-inflammatory cytokines such as Interleukin 6, Tumor Necrosis Factor- α and C- reactive protein. In the present review, an attempt has been made to elucidate the molecular mechanisms involved in xenoestrogen-induced obesity and metainflammation. Further the role of immune system and the involvement of metabolism in the genesis of metainflammation have also been explored briefly.

Introduction

According to the World Heart Federation (WHF) report of 2022, approximately 2.3 billion individuals were suffering due to obesity worldwide. Interestingly, the number of obese people is higher as compared to the underweight in almost every part of the world. As per the currently ongoing trend, approximately 2.7 billion adults i.e. around 33% of the total global population are supposed to be obese by the year 2025.¹ Along with obesity, the cases of other metabolic diseases like type 2 diabetes mellitus and non-alcoholic fatty liver disease (NAFLD) are also rising significantly.^{2,3} Therefore, scientists are exploring to find out the intricate relationship among different metabolic diseases as well as to understand the pattern of disease initiation and subsequent complications. In order to decipher this relationship, the research in the last decades has been oriented towards the concept of immunometabolism, an interface between metabolism and immune system. Apparently, it seems that nutrient- (metabolic process) and pathogen-sensing (immune system) systems are working in a mutually exclusive manner

Keywords: Metainflammation, Obesity, Xenoestrogen, Bisphenol A, Macrophage, Immunometabolism in higher organisms. In lower invertebrates such as drosophila, these two systems are not only working intricately but also are located intertwiningly, known as the fat body.⁴ The close association between these two consistently working systems gave birth to a new concept called "metainflammation", the term which was first coined by Hotamisligil in year 2006.⁵ In most of obese people, the proliferating adipocytes initiate a cascade of low grade systemic inflammation known as metainflammation. If this persists for a longer period, it can propagate deleterious effects to the metabolically active organs such as pancreas, liver and heart.^{6,3}

It is undeniable that unhealthy dietary practices and a sedentary lifestyle have been considered as the predominant contributing factors in continuously surging obesity cases. Moreover, an emerging body of scientific literature emphasizes that exposure to different environmental chemicals act in tandem with these factors to exacerbate the incidence of obesity at an alarming degree.⁷ For instance, xenoestrogens, a group of chemicals which mimic the natural estrogen synthesized in the body, play a pivotal role in the development of obesity, other metabolic diseases and metainflammation.^{8,9} Among the xenoestrogens, BPA is the most abundant and most studied chemical in both humans and experimental

Review Article

models.^{10,11} Various *in-vivo* studies have shown that pre- and post-natal exposure to Bisphenol A (BPA) increased the body weight of pups and induced permanent obesity in the adult rodents.¹²⁻¹⁴ Therefore, it is clear that these xenoestrogens can target the early phases of life leading to the lifelong effects. Being non-biodegradable and highly lipophilic in nature, the xenoestrogens tend to accumulate in the adipose tissue and cause inflammation for a longer period.¹⁵ In the present article, an attempt has been made to provide a critical insight regarding the role of metainflammation in xenoestrogeninduced obesity. Further, the molecular aspects of the existing interface between xenoestrogens, immune system and genesis of metainflammation has also been explored briefly.

Role of Xenoestrogens in Obesity

Xenoestrogen can cause obesity mainly through metabolic disruption.^{16,17} The obesogenic actions of BPA have been extensively studied in the in-vitro and *in-vivo* models.¹⁶ Ultimately, the mechanisms through which these xenoestrogens exert their metabolic disruption and obesogenic effect, even when the level of exposure is below the No Observed Adverse Effect Level (NOAEL), are being elucidated by different scientific studies. The relationship between the exposure level of xenoestrogens and weight gain forms an inverted "U" shape, curve where the lower exposure level exerts more deleterious effect than the higher exposure level.¹⁸ Pro-opiomelanocortin (POMC) neurons in the hypothalamus restrict food intake while Agouti-Related Peptide (AgRP) and Neuropeptide Y (NPY) do the reverse effect. The experimental in-vivo models showed that BPA-exposed mice had POMC neuron innervations and the AgRP and NPY peptides activities were abolished.^{19,20} Apart from doing dysbalance in neuronal control, xenoestrogens can alter the carbohydrate metabolism by causing insulin resistance and necrosis of pancreatic β -cells.²¹ They also increase the number as well as the size of white adipocytes by elevating triglyceride content, lipoprotein lipase activity and adipogenic transcription factor expression like CCAAT Enhancer Binding Protein-ß (C/EBP-ß).^{7,22} Thus, xenoestrogens perturb the intricate balance between the neuronal control and metabolism of the body and cause excess weight gain as well as obesity.¹⁶

Xenoestrogens as a Causative Factor for Metainflammation

Xenoestrogens are compounds which have an ability to interfere with the natural hormones which are accountable for the regulation of various physiological functions such as development,

behavior, fertility as well as maintaining the homeostasis in the body.²³ The matter of concern is that there are several xenoestrogens which are lipophilic in nature. As Body Mass Index (BMI) increases, these lipophilic molecules accumulate in the adipose tissue. They are non-biodegradable; therefore their concentration tends to increase with time and further continuous exposure is a subject of concern.²⁴ Basically, there are two types of xenoestrogens: natural (phytoestrogens) and synthetic²³, however some authors do not consider natural ones in the list of xenoestrogens.^{25,26} Among the synthetic xenoestrogens, BPA is the most abundant in use; whereas isoflavones form the most important group of phytoestrogens.²⁷ Although, isoflavones exhibit antioxidant, anticancer, antimicrobial and anti-inflammatory properties, some of them like soy isoflavones are also reported to cause obesity and other metabolic disorders.^{28,29} Both of them imitate the action of estrogen and alter the metabolic processes in the body. These are capable to upregulate different transcription factors like Peroxisome Proliferator-Activated Receptor (PPARγ), C/EBP and Nuclear Factor Erythroid 2-related Factor 2 (Nrf2), which can ultimately lead to the induction of obesity and associated metainflammation.¹⁴ Table 1 depicts some of the important xenoestrogens, their sources along with their role in obesity and metainflammation.

Many of xenoestrogens cause metainflammation in a similar way like BPA does. BPA significantly increased the expression of several genes involved in adipogenesis and lipid accumulation, including C/ EBP α , C/EBP β , PPAR γ , Fatty Acid Synthetase (FASN) and Sterol Regulatory Element Binding Protein1c (SREBP1c). The up-regulation of these adipogenic transcription factors and enzymes are reported to be involved in the development of metainflammation. ³⁰⁻³² BPA has an ability to interact with Nuclear Receptors (NR) including Retinoid X Receptor (RXR), PPAR γ , Estrogen Receptors (ER), Thyroid Receptors (TR) and Glucocorticoid Receptors (GR), as a result of which it can induce differentiation of adipocytes and lipid accumulation.³³ BPA can also activate classical transduction pathways of $ER\alpha$ and $ER\beta$, which can further reduce adiponectin secretion and increase the proliferation of adipocytes.³⁴ Organochlorines like Dichlorodiphenyltrichloroethane (DDT) and endosulfan increase the expression of Aryl Hydrocarbon Receptor (AHR) transcription factor, which has the potential to elevate the production of the aromatase, a CYP450 mediated enzyme.^{35,36} Aromatase, in turn, converts androgen to estrogen which upregulates the PPAR and c/EBP transcription factors.³⁶ PPAR and ER family has been reported to correlate the different molecular

pathways involved in the process of metainflammation.⁵ Males are more prone to metainflammation and related metabolic diseases as compared to age-matched pre-menopausal females.³⁷ This type of difference in susceptibility is due to the presence of estrogen, which plays a protective role in pre-menopausal females. Further, post-menopausal females develop more metabolic disorders than age-matched males.³⁸

Metainflammation and Immune System: The Connecting Link

Among the immune cells, macrophages significantly contribute to the obesity-induced systemic inflammation. Macrophages have an ability to quickly sense their microenvironment and change their metabolic profile as well as express a wide variety of inflammatory markers.⁴⁴ Under the stressful conditions, macrophages are polarized into proinflammatory macrophages (M1 subtype), which mainly secretes pro-inflammatory cytokines. Once the stress is over, another type of macrophages (M2 subtype) get activated and play a crucial role in tissue repairing.⁴⁵ Interestingly, M2 macrophages utilize fatty acids as the energy source and produce ATP through β -oxidation and oxidative phosphorylation, which is more time consuming. Conversely, M1 macrophages quickly undergo aerobic glycolysis.⁴⁶ Macrophages during metainflammation reportedly behave like M1 macrophages to a great extent.47 These metabolically activated macrophages express low levels of CD206 (overexpressed in macrophages of non-obese animals) and elevated the levels of CD11c, CD36, Macrophage Scavenger Receptor 1 (MSR1), ATPbinding cassette A1 (ABCA1), adipose differentiationrelated protein such as Perilipin-2.48,46,49 Various types of immune cells in the adipose tissue can affect the shift in macrophage polarization. For instance, neutrophils induce this change by using protease elastase, T-lymphocytes by using interferon-y, natural killer cells induce polarization by TNF- α and MCP1 and B cells contribute by producing IgG antibodies.^{50,47} Macrophages expressing CD11c have been linked to insulin resistance and are located in the crown-like structures, which encircle necrotic adipocytes to eliminate them through a process called exophagy. ⁴⁶ This process results in the uptake of FFA and lipids by macrophages and the formation of foam cells.⁵¹ In summary, obesity induces some changes in the phenotype and behaviour of macrophages which contribute greatly to the overactivation of innate immunity system of our body leading to the initiation of low grade systemic inflammation.

Obesity and Metainflammation: The Molecular Basis

Lipids are involved in the coordinated regulation of metabolic, inflammatory and innate immune processes. The quest for elucidating the molecular signaling involved in metainflammation started around a decade ago when researchers discovered high levels of Tumor Necrosis Factor- α (TNF- α) in the adipose tissue of obese mice. This finding established a clear connection between obesity and chronic inflammation in the experimental mice model.⁵² Later, it was established that TNF- α is not released by adipocytes but rather by the macrophages which surround them.⁵³ TNF- α is a marker of local as well as systemic inflammation, therefore high chances of crosstalk existed between adipocytes and immune system under obese condition. 54,55 According to literature, Endoplasmic Reticulum (ER) can serve as a common target and in fact, it is considered that ER begins the inflammatory cascade in the metainflammation process.5,56

In obesity, the elevated levels of Free Fatty Acid (FFA) in the adipocytes increase ER stress which can further activate a number of inter-connected pathways.⁵⁷ FFA stimulates the unfolded protein response which is mediated by nutrient fluctuations, hypoxia and the presence of different toxins. This, in turn, generates additional molecular targets such as Activating Transcription Factor 6 (ATF-6), Inositol-Requiring Enzyme Type 1 (IRE1) and Protein Kinase R-like Endoplasmic Reticulum Kinase (PERK), which can further lead to metainflammation.^{58,59} It has been reported that ATF-6 increases stress to Golgi body, IRE1 stimulates the lipid droplet formation and PERK binds with PPAR and C/EBP proteins.⁶⁰ These three phenomena can increase the oxidative stress in the system and activate Jun N-terminal Kinase (JNK) and IkB Kinase Complex (IKK) factors to cause inflammation and necrosis of adipocytes.⁶¹ Additionally, FFA directly upregulates the expression of PPAR, C/EBP and Nrf2 as depicted in the figure 1. All of these mediators alter the expression of IL-6 and TNF- α which can further stimulate monocyte to macrophage activation. The macrophages also produce IL-6 and TNF- α which also help to initiate the vicious cycle of inflammation.⁵⁹

The leakage of calcium (Ca²⁺) ions from the outer membrane of ER is another phenomenon which takes place under ER stress. The excess of Ca²⁺ ions are responsible for the mitochondrial damage and secretion of cytochrome-c. This further leads to apoptosis by binding with the Apoptotic Protease Activating Factor-1 (Apaf-1).⁶² Both ER stress and mitochondrial damage increase the Reactive Oxygen

Review Article

Species (ROS) levels which in turn activate JNK and IKK. Both JNK and IKK up-regulate the Monocyte Chemoattractant Protein-1 (MCP-1), IL6 and TNF- α expression but decrease the production of adiponectin, a cytokine produced exclusively by the adipocytes.⁶³ Reduction in the adiponectin level is also accomplished by GPR30, a G-protein coupled receptor involved in the production of antiinflammatory cytokines like IL10. Estrogen can have a rapid non-genomic response via GPR30. The GPR30 knockout mice have reportedly exhibited elevated levels of pro-inflammatory cytokines and low adiponectin levels in their circulation.⁶⁴ The reduced production of adiponectin and increased production of pro-inflammatory cytokines block the action of Insulin Receptor Substrate (IRS), resulting in the insulin resistance in adipocytes. Insulin receptor via mTORC1-Egr1-ATGL pathway ameliorates the degradation of triglyceride into FFA which increases the size of lipid droplet.⁶⁵ Due to large lipid droplets, ER tends to synthesize more proteins to package the enlarged lipid droplets and this phenomenon is responsible for the excessive ER stress.⁶⁶ From the existing literature, it is clearly evident that the pathophysiology of metainflammation relies on a complex intertwined pathways involving the metabolic and immune system, which start with ER stress and then progress to other cell-organelles leading to the production of pro-inflammatory cytokines. The purinergic system, specifically the metabolites ATP and adenosine, plays a significant role in the development of metainflammation.⁶⁷ Adenosine exhibits anti-inflammatory properties by inhibiting Th1-polarizing responses and promoting the production of anti-inflammatory cytokines and Th2-polarizing responses. On the other hand, ATP, particularly at high extracellular concentrations, contributes to inflammation and cell death.68,69 In metainflammation, there is a decrease in adenosine levels and a significant increase in ATP levels.⁶⁹

Metabolic Considerations in Metainflammation: Role of Pancreas and Liver

In the context of metainflammation, the pancreas and liver are primarily affected. In the pancreas, metainflammation is associated with two main inflammatory pathways, JNK-AP-1 and IKK-NF- κ B, which are connected to IRE-1 and PERK activity during ER stress.⁷⁰ These pathways involve interactions between IRE-1 and JNK activation through TNF receptor-associated factor 2 (TRAF2), as well as the association of IRE-1 and PERK activation with the IKK-NF- κ B pathway. The activation of IRE-1 and PERK are also associated with the IKK-NF- κ B pathway, but through distinct mechanisms. IRE-1 interacts with IKK- β through

TRAF2, whereas PERK activation leads to the degradation of I κ B, thereby facilitating NF- κ B activity.^{71,72} In the liver, metainflammation occurs due to the entry of excessive amounts of free fatty acids (FFA) from necrotic adipose tissue, leading to lipid accumulation in hepatocytes and induce lipotoxicity.^{73,74} The liver, like adipose tissue, has resident macrophages called Kupffer cells, and the interaction between hepatocytes and Kupffer cells follows a similar pattern as in adipose tissue. As the liver is crucial for carbohydrate and fat metabolism and relies on insulin, the excess lipid accumulation results in insulin resistance and the production of pro-inflammatory cytokines, disrupting the metabolic regulation.^{75,76}

Future perspectives

The purinergic system is a crucial modulator of metainflammation and its role has been investigated in the aetiology of osteoarthritis.⁶⁷ The exploration of the role of purinergic system mediated metainflammation in metabolic disorders might open up a new therapeutic avenue for the disease. The relationship between metabolism and inflammation via epigenetic regulation of gene expression is another area which needs more research and may eventually lead to a potential clinical intervention strategy. Recent research suggests that lysine acetylation of both histone and non-histone proteins cause alterations in energy metabolism during chronic inflammation.⁷⁷ Therefore, by regulating the expression of pro- and anti-inflammatory mediators, deacetylase inhibitors or activators may be further strategies to prevent macrophage-induced metainflammation. Despite the significant progress made over the past few decades, many concerns regarding the processes of macrophage polarisation and metainflammation remains unanswered. The interaction between macrophages and their milieu is a complicated and dynamic process due to the heterogeneous and versatile character of macrophages. At present the understanding of these interactions in *in-vivo* conditions are limited. These issues can be addressed through the incorporation of new technology, such as computational biological methods for better understanding and interpretation.

References

- 1. Federation WH. Obesity. https://world-heartfederation.org/what-we-do/obesity/. 2022, Accessed July 05, 2023.
- Yang M, Liu S, Zhang C. The Related Metabolic Diseases and Treatments of Obesity, Healthcare (Basel), 10 (2022) 10.3390/healthcare10091616.
- Valenzuela PL, Carrera-Bastos P, Castillo-García A, Lieberman DE, Santos-Lozano A, Lucia A. Obesity and the risk of cardiometabolic diseases, Nat Rev Cardiol,

Review Article

20 (2023) 475-94. 10.1038/s41569-023-00847-5.

- Rusten TE, Lindmo K, Juhász G, et al. Programmed autophagy in the Drosophila fat body is induced by ecdysone through regulation of the PI3K pathway, Dev Cell, 7 (2004) 179-92. 10.1016/j.devcel.2004.07.005.
- 5. Hotamisligil GS. Inflammation and metabolic disorders, Nature, 444 (2006) 860-7. 10.1038/nature05485.
- Cao H. Adipocytokines in obesity and metabolic disease, J Endocrinol, 220 (2014) T47-59. 10.1530/joe-13-0339.
- Salehpour A, Shidfar F, Hedayati M, Neshatbini Tehrani A, Farshad AA, Mohammadi S. Bisphenol A enhances adipogenic signaling pathways in human mesenchymal stem cells, Genes Environ, 42 (2020) 13. 10.1186/ s41021-020-00150-6.
- Wang LH, Chen LR, Chen KH. In Vitro and Vivo Identification, Metabolism and Action of Xenoestrogens: An Overview, Int J Mol Sci, 22 (2021) 10.3390/ ijms22084013.
- 9. Kumar M, Sarma DK, Shubham S, et al. Environmental Endocrine-Disrupting Chemical Exposure: Role in Non-Communicable Diseases, Front Public Health, 8 (2020) 553850. 10.3389/fpubh.2020.553850.
- Wu W, Li M, Liu A, et al. Bisphenol A and the Risk of Obesity a Systematic Review With Meta-Analysis of the Epidemiological Evidence, Dose-Response, 18 (2020) 1559325820916949. 10.1177/1559325820916949.
- Moon S, Seo MY, Choi K, Chang YS, Kim SH, Park MJ. Urinary bisphenol A concentrations and the risk of obesity in Korean adults, Sci Rep, 11 (2021) 1603. 10.1038/s41598-021-80980-8.
- Zhou Z, Lei Y, Wei W, et al. Association between prenatal exposure to bisphenol a and birth outcomes: A systematic review with meta-analysis, Medicine (Baltimore), 98 (2019) e17672. 10.1097/ md.00000000017672.
- Newbold RR, Padilla-Banks E, Jefferson WN. Environmental estrogens and obesity, Mol Cell Endocrinol, 304 (2009) 84-9. 10.1016/ j.mce.2009.02.024.
- 14. Cimmino I, Fiory F, Perruolo G, et al. Potential Mechanisms of Bisphenol A (BPA) Contributing to Human Disease, Int J Mol Sci, 21 (2020) 10.3390/ ijms21165761.
- 15. La Merrill MA, Vandenberg LN, Smith MT, et al. Consensus on the key characteristics of endocrinedisrupting chemicals as a basis for hazard identification, Nat Rev Endocrinol, 16 (2020) 45-57. 10.1038/s41574-019-0273-8.
- Oliviero F, Marmugi A, Viguié C, Gayrard V, Picard-Hagen N, Mselli-Lakhal L. Are BPA Substitutes as Obesogenic as BPA?, Int J Mol Sci, 23 (2022) 10.3390/ ijms23084238.
- Shahnazaryan U, Wójcik M, Bednarczuk T, Kurylowicz A. Role of Obesogens in the Pathogenesis of Obesity, Medicina (Kaunas), 55 (2019) 10.3390/ medicina55090515.
- Angle BM, Do RP, Ponzi D, et al. Metabolic disruption in male mice due to fetal exposure to low but not high doses of bisphenol A (BPA): evidence for effects on body weight, food intake, adipocytes, leptin, adiponectin, insulin and glucose regulation, Reprod Toxicol, 42 (2013) 256-68. 10.1016/ j.reprotox.2013.07.017.
- 19. MacKay H, Patterson ZR, Abizaid A. Perinatal Exposure to Low-Dose Bisphenol-A Disrupts the Structural and Functional Development of the Hypothalamic Feeding Circuitry, Endocrinology, 158 (2017) 768-77. 10.1210/ en.2016-1718.

- Salehi A, Loganathan N, Belsham DD. Bisphenol A induces Pomc gene expression through neuroinflammatory and PPAR? nuclear receptormediated mechanisms in POMC-expressing hypothalamic neuronal models, Mol Cell Endocrinol, 479 (2019) 12-19. 10.1016/j.mce.2018.08.009.
- Martinez-Pinna J, Marroqui L, Hmadcha A, et al. Oestrogen receptor β mediates the actions of bisphenol-A on ion channel expression in mouse pancreatic beta cells, Diabetologia, 62 (2019) 1667-80. 10.1007/ s00125-019-4925-y.
- 22. Choi SI, Kwon HY, Han X, et al. Environmental obesogens (bisphenols, phthalates and parabens) and their impacts on adipogenic transcription factors in the absence of dexamethasone in 3T3-L1 cells, J Steroid Biochem Mol Biol, 214 (2021) 105994. 10.1016/ j.jsbmb.2021.105994.
- 23. Paterni I, Granchi C, Minutolo F. Risks and benefits related to alimentary exposure to xenoestrogens, Crit Rev Food Sci Nutr, 57 (2017) 3384-404. 10.1080/ 10408398.2015.1126547.
- Le Magueresse-Battistoni B. Adipose Tissue and Endocrine-Disrupting Chemicals: Does Sex Matter?, Int J Environ Res Public Health, 17 (2020) 10.3390/ ijerph17249403.
- Wang X, Ha D, Yoshitake R, Chan YS, Sadava D, Chen S. Exploring the Biological Activity and Mechanism of Xenoestrogens and Phytoestrogens in Cancers: Emerging Methods and Concepts, Int J Mol Sci, 22 (2021) 10.3390/ijms22168798.
- Wiseman A, Goldfarb PS, Ridgway TJ, Wiseman H. Xenoestrogens and phytoestrogens: bioremediation or biomonitoring?, Trends Biotechnol, 17 (1999) 43. 10.1016/s0167-7799(98)01255-4.
- Adetunji CO, Akram M, Mtewa AG, et al., Chapter 18 -Biochemical and pharmacotherapeutic potentials of lycopene in drug discovery In: C. Egbuna, A.P. Mishra and M.R. Goyal, Editors, Preparation of Phytopharmaceuticals for the Management of Disorders, Academic Press (2021), pp. 307-60.
- Desmawati D, Sulastri D. Phytoestrogens and Their Health Effect, Open Access Maced J Med Sci, 7 (2019) 495-99. 10.3889/oamjms.2019.044.
- Akhlaghi M, Zare M, Nouripour F. Effect of Soy and Soy Isoflavones on Obesity-Related Anthropometric Measures: A Systematic Review and Meta-analysis of Randomized Controlled Clinical Trials, Advances in Nutrition, 8 (2017) 705-17. https://doi.org/10.3945/ an.117.015370.
- Rahman SM, Janssen RC, Choudhury M, et al. CCAAT/ Enhancer-binding Protein ß (C/EBPß) Expression Regulates Dietary-induced Inflammation in Macrophages and Adipose Tissue in Mice*, Journal of Biological Chemistry, 287 (2012) 34349-60. https://doi.org/ 10.1074/jbc.M112.410613.
- 31. Wei X, Song H, Yin L, et al. Fatty acid synthesis configures the plasma membrane for inflammation in diabetes, Nature, 539 (2016) 294-98. 10.1038/nature20117.
- Kolehmainen M, Vidal H, Alhava E, Uusitupa MI. Sterol regulatory element binding protein 1c (SREBP-1c) expression in human obesity, Obes Res, 9 (2001) 706-12. 10.1038/oby.2001.95.
- Ahmed S, Atlas E. Bisphenol S- and bisphenol A-induced adipogenesis of murine preadipocytes occurs through direct peroxisome proliferator-activated receptor gamma activation, Int J Obes (Lond), 40 (2016) 1566-73. 10.1038/ijo.2016.95.
- 34. Choi B-I, Harvey AJ, Green MP. Bisphenol A affects early bovine embryo development and metabolism that is

<u>Review Article</u>

negated by an oestrogen receptor inhibitor, Scientific Reports, 6 (2016) 29318. 10.1038/srep29318.

- 35. Darbre PD. Endocrine Disruptors and Obesity, Curr Obes Rep, 6 (2017) 18-27. 10.1007/s13679-017-0240-4.
- Pestana D, Teixeira D, Meireles M, et al. Adipose tissue dysfunction as a central mechanism leading to dysmetabolic obesity triggered by chronic exposure to p,p'-DDE, Sci Rep, 7 (2017) 2738. 10.1038/s41598-017-02885-9.
- Varghese M, Griffin C, Abrishami S, et al. Sex hormones regulate metainflammation in diet-induced obesity in mice, J Biol Chem, 297 (2021) 101229. 10.1016/ j.jbc.2021.101229.
- Ko SH, Kim HS. Menopause-Associated Lipid Metabolic Disorders and Foods Beneficial for Postmenopausal Women, Nutrients, 12 (2020) 10.3390/nu12010202.
- De Coster S, van Larebeke N. Endocrine-disrupting chemicals: associated disorders and mechanisms of action, J Environ Public Health, 2012 (2012) 713696. 10.1155/2012/713696.
- Søgaard R, Holbech H, Nørum U, Bjerregaard P. Investigation of the in vivo estrogenicity of the UV-filters 4-methylbenzylidene camphor and octyl methoxy cinnamate in rainbow trout (Oncorhynchus mykiss), Ecotoxicology and Environmental Safety, 224 (2021) 112657. https://doi.org/10.1016/ j.ecoenv.2021.112657.
- 41. Marshall BL, Liu Y, Farrington MJ, et al. Early genistein exposure of California mice and effects on the gut microbiota-brain axis, J Endocrinol, 242 (2019) 139-57. 10.1530/joe-19-0214.
- 42. White R, Jobling S, Hoare SA, Sumpter JP, Parker MG. Environmentally persistent alkylphenolic compounds are estrogenic, Endocrinology, 135 (1994) 175-82. 10.1210/en.135.1.175.
- Shin J, Lee E, An S, et al. Benzophenone-3 and benzophenone-8 exhibit obesogenic activity via peroxisome proliferator-activated receptor ? pathway, Toxicology in Vitro, 67 (2020) 104886. 10.1016/ j.tiv.2020.104886.
- 44. Parisi L, Gini E, Baci D, et al. Macrophage Polarization in Chronic Inflammatory Diseases: Killers or Builders?, J Immunol Res, 2018 (2018) 8917804. 10.1155/2018/ 8917804.
- 45. Talukdar S, Oh DY, Bandyopadhyay G, et al. Neutrophils mediate insulin resistance in mice fed a high-fat diet through secreted elastase, Nat Med, 18 (2012) 1407-12. 10.1038/nm.2885.
- Russo S, Kwiatkowski M, Govorukhina N, Bischoff R, Melgert BN. Meta-Inflammation and Metabolic Reprogramming of Macrophages in Diabetes and Obesity: The Importance of Metabolites, Front Immunol, 12 (2021) 746151. 10.3389/fimmu.2021.746151.
- 47. Castoldi A, Naffah de Souza C, Câmara NO, Moraes-Vieira PM. The Macrophage Switch in Obesity Development, Front Immunol, 6 (2015) 637. 10.3389/ fimmu.2015.00637.
- 48. Huang SC, Everts B, Ivanova Y, et al. Cell-intrinsic lysosomal lipolysis is essential for alternative activation of macrophages, Nat Immunol, 15 (2014) 846-55. 10.1038/ni.2956.
- 49. Kratz M, Coats Brittney R, Hisert Katherine B, et al. Metabolic Dysfunction Drives a Mechanistically Distinct Proinflammatory Phenotype in Adipose Tissue Macrophages, Cell Metabolism, 20 (2014) 614-25. https://doi.org/10.1016/j.cmet.2014.08.010.
- 50. Lee BC, Kim MS, Pae M, et al. Adipose Natural Killer Cells Regulate Adipose Tissue Macrophages to Promote

Insulin Resistance in Obesity, Cell Metab, 23 (2016) 685-98. 10.1016/j.cmet.2016.03.002.

- Haka AS, Barbosa-Lorenzi VC, Lee HJ, et al. Exocytosis of macrophage lysosomes leads to digestion of apoptotic adipocytes and foam cell formation[S], Journal of Lipid Research, 57 (2016) 980-92. https://doi.org/10.1194/ jlr.M064089.
- Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance, Science, 259 (1993) 87-91. 10.1126/science.7678183.
- 53. Surmi BK, Hasty AH. Macrophage infiltration into adipose tissue: initiation, propagation and remodeling, Future Lipidol, 3 (2008) 545-56. 10.2217/17460875.3.5.545.
- 54. Sethi JK, Hotamisligil GS. Metabolic Messengers: tumour necrosis factor, Nat Metab, 3 (2021) 1302-12. 10.1038/ s42255-021-00470-z.
- 55. Hill AA, Reid Bolus W, Hasty AH. A decade of progress in adipose tissue macrophage biology, Immunol Rev, 262 (2014) 134-52. 10.1111/imr.12216.
- Fernandes-da-Silva A, Miranda CS, Santana-Oliveira DA, et al. Endoplasmic reticulum stress as the basis of obesity and metabolic diseases: focus on adipose tissue, liver, and pancreas, Eur J Nutr, 60 (2021) 2949-60. 10.1007/ s00394-021-02542-y.
- 57. Acosta-Martinez M, Cabail MZ. The PI3K/Akt Pathway in Meta-Inflammation, Int J Mol Sci, 23 (2022) 10.3390/ ijms232315330.
- Siwecka N, Rozpedek-Kaminska W, Wawrzynkiewicz A, Pytel D, Diehl JA, Majsterek I. The Structure, Activation and Signaling of IRE1 and Its Role in Determining Cell Fate, Biomedicines, 9 (2021) 10.3390/ biomedicines9020156.
- 59. Zha BS, Zhou H. ER Stress and Lipid Metabolism in Adipocytes, Biochem Res Int, 2012 (2012) 312943. 10.1155/2012/312943.
- Coll T, Barroso E, Alvarez-Guardia D, et al. The Role of Peroxisome Proliferator-Activated Receptor beta/delta on the Inflammatory Basis of Metabolic Disease, PPAR Res, 2010 (2010) 10.1155/2010/368467.
- 61. Yung JHM, Giacca A. Role of c-Jun N-terminal Kinase (JNK) in Obesity and Type 2 Diabetes, Cells, 9 (2020) 10.3390/cells9030706.
- Eleftheriadis T, Pissas G, Liakopoulos V, Stefanidis I. Cytochrome c as a Potentially Clinical Useful Marker of Mitochondrial and Cellular Damage, Front Immunol, 7 (2016) 279. 10.3389/fimmu.2016.00279.
- Ghantous CM, Azrak Z, Hanache S, Abou-Kheir W, Zeidan A. Differential Role of Leptin and Adiponectin in Cardiovascular System, Int J Endocrinol, 2015 (2015) 534320. 10.1155/2015/534320.
- 64. Sharma G, Prossnitz ER. Targeting the G protein-coupled estrogen receptor (GPER) in obesity and diabetes, Endocr Metab Sci, 2 (2021) 10.1016/ j.endmts.2021.100080.
- 65. Kim OK, Jun W, Lee J. Mechanism of ER Stress and Inflammation for Hepatic Insulin Resistance in Obesity, Ann Nutr Metab, 67 (2015) 218-27. 10.1159/000440905.
- 66. Krahmer N, Farese RV, Jr., Walther TC. Balancing the fat: lipid droplets and human disease, EMBO Mol Med, 5 (2013) 973-83. 10.1002/emmm.201100671.
- Gratal P, Lamuedra A, Medina JP, et al. Purinergic System Signaling in Metainflammation-Associated Osteoarthritis, Front Med (Lausanne), 7 (2020) 506. 10.3389/ fmed.2020.00506.
- 68. Cauwels A, Rogge E, Vandendriessche B, Shiva S, Brouckaert P. Extracellular ATP drives systemic inflammation, tissue damage and mortality, Cell Death

& Disease, 5 (2014) e1102-e02. 10.1038/ cddis.2014.70.

- 69. Trautmann A. Extracellular ATP in the immune system: more than just a "danger signal", Sci Signal, 2 (2009) pe6. 10.1126/scisignal.256pe6.
- Li M, Chi X, Wang Y, Setrerrahmane S, Xie W, Xu H. Trends in insulin resistance: insights into mechanisms and therapeutic strategy, Signal Transduct Target Ther, 7 (2022) 216. 10.1038/s41392-022-01073-0.
- Chen L, Chen R, Wang H, Liang F. Mechanisms Linking Inflammation to Insulin Resistance, Int J Endocrinol, 2015 (2015) 508409. 10.1155/2015/508409.
- 72. Wen X, Zhang B, Wu B, et al. Signaling pathways in obesity: mechanisms and therapeutic interventions, Signal Transduct Target Ther, 7 (2022) 298. 10.1038/ s41392-022-01149-x.
- Arslan N. Obesity, fatty liver disease and intestinal microbiota, World J Gastroenterol, 20 (2014) 16452-63. 10.3748/wjg.v20.i44.16452.

- 74. Godoy-Matos AF, Silva Júnior WS, Valerio CM. NAFLD as a continuum: from obesity to metabolic syndrome and diabetes, Diabetology & Metabolic Syndrome, 12 (2020) 60. 10.1186/s13098-020-00570-y.
- Glen J, Floros L, Day C, Pryke R. Non-alcoholic fatty liver disease (NAFLD): summary of NICE guidance, Bmj, 354 (2016) i4428. 10.1136/bmj.i4428.
- Fabbrini E, Sullivan S, Klein S. Obesity and nonalcoholic fatty liver disease: biochemical, metabolic, and clinical implications, Hepatology, 51 (2010) 679-89. 10.1002/ hep.23280.
- Drazic A, Myklebust LM, Ree R, Arnesen T. The world of protein acetylation, Biochim Biophys Acta, 1864 (2016) 1372-401. 10.1016/j.bbapap.2016.06.007.

International Conference cum Workshop on ARTIFICIAL INTELLIGENCE SOLUTIONS FOR PHARMACEUTICAL RESEARCH AND KNOWLEDGE (AI-SPARK 2023)

October 9-11, 2023

Organized by DEPARTMENT OF PHARMACOINFORMATICS NIPER, S.A.S. Nagar, Punjab-160062

ABOUT THE CONFERENCE

AI-SPARK 2023 aims to bring together the leading academicians, industrialists, researchers, and scholars across the globe to facilitate a better understanding and integration of the latest artificial intelligence approaches and propel the development of new tools and techniques in drug design and development.

HANDS ON TRAINING

Two days workshop (Oct. 10-11, 2023) will cover applications of machine leaning and artificial intelligence using python programming. Due to the limited availability of computer systems, participants will be considered on a first-comefirst-served basis.

CONFERENCE TRACKS

- AI in Drug Discovery
- AI in Protein Structure Prediction/Omics
- AI in Pharmacokinetics and Toxicity Predictions
- AI in Drug Delivery and Formulations
- AI in Medical Devices
- AI in Translational Research
- Ethical, legal, and regulatory considerations in AI-driven drug discovery
- Poster Session
- Hands on AI/ML training with python

IMPORTANT DATES	REGISTRATION		CONTACT
Conference: Oct. 9-11, 2023	Category	Registration	https://niper.gov.in/aispark
Workshop: Oct 10-11 2023	Ctudopto/	Fees (INR)	Registration:
Workshop. Occ. 10-11, 2025	Students/	2000	https://forms.gle/
Last Date Registration: Sept. 15, 2023	Research Scholars	2000	59/UCPYZ/FL64/5N9
	Faculty	5000	Email:
Poster Submission: Sept. 15, 2023	Industry		aispark.niper@gmail.com,
	Professionals	10000	pi@niper.ac.in

SPONSORED BY

NIPER, S.A.S. Nagar; Department of Biotechnology (DBT), Government of India